

# GoldiBlot™ His Western Blot Kit



95 Horseblock Road, Unit 1, Yaphank NY 11980-9710  
Tel: (877) 447-6266 (Toll-Free in US) or (631) 205-9490 Fax: (631) 205-9493  
Tech Support: (631) 205-9492 [tech@nanoprobes.com](mailto:tech@nanoprobes.com)  
[www.nanoprobes.com](http://www.nanoprobes.com)

## PRODUCT INFORMATION

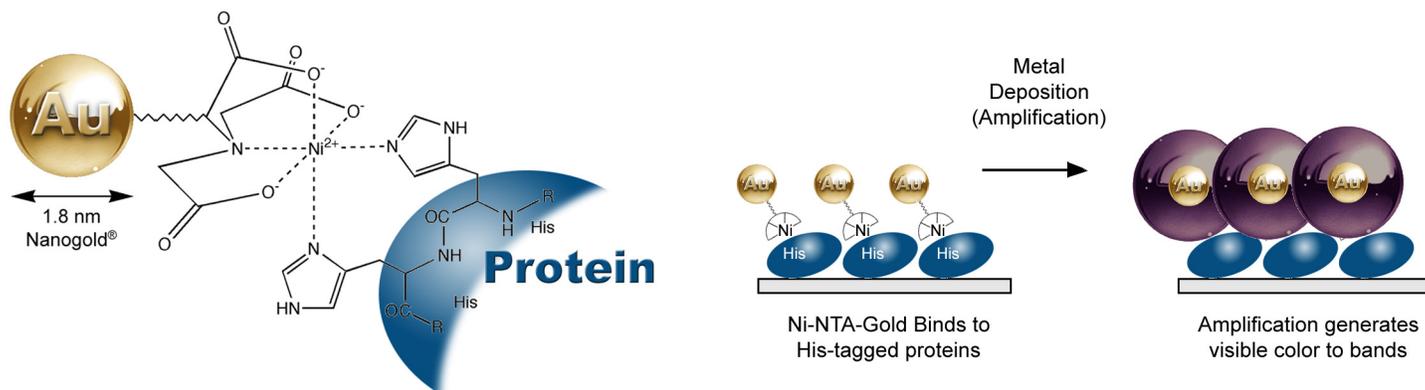
**Product Name:** GoldiBlot™ His Western Blot Kit  
**Catalog Number:** 2090-10BLOTS  
**Revision:** 1.6 (February 2024)

### INTENDED USE

GoldiBlot™ His Western Blot Kit is intended and optimized for direct visualization of recombinant His-tagged proteins and other proteins bearing different polyhistidine (His) tags in western or dot blotting applications. Unlike detection by some anti-6xHis antibodies, GoldiBlot™ His detection does not require a specific location of the polyhistidine tag (*N*- or *C*-terminus) or the presence of specific adjacent amino acid sequences, and can recognize internal His-tags in addition to those at the *N*- or *C*-terminus. Probing is complete in one step, without the need for primary and secondary antibody incubations.

### PRINCIPLE OF GOLDIBLOT™ HIS WESTERN BLOT KIT

GoldiBlot™ His Western Blot Kit uses Ni-NTA (nickel-nitrilotriacetic acid)-functionalized gold nanoparticles to specifically bind to His-tagged proteins<sup>1-6</sup>. When autometallographic amplification is subsequently applied to the gold nanoparticles, GoldiBlot™ allows the direct visualization of His-tagged proteins. GoldiBlot™ generates specific purple-colored metallic bands or dots which do not fade and will not dissolve in water and organic solvents. The GoldiBlot™ His Western Blot Kit can detect nanogram levels of purified His-tagged proteins. The entire procedure takes about 1 hour.



*(left)* Ni-NTA-Gold, showing mechanism of binding to a polyhistidine (His) – tagged protein. *(right)* Principle of GoldiBlot™: gold binding followed by autometallographic amplification (deposition of metal selectively onto the gold particles) generates visible signal.

**REAGENTS PROVIDED**

#2090-10BLOTS kit contains the following materials:

GoldiBlot™ Nickel-NTA-Au	1 mL
GoldiBlot™ AutoMet Detect A	27 mL
GoldiBlot™ AutoMet Detect B	27 mL
GoldiBlot™ AutoMet Detect C	27 mL
GoldiBlot™ AutoMet Detect D	27 mL

**MATERIALS REQUIRED, BUT NOT SUPPLIED**

**TBS-0.1%T:** 20 mM Tris, 0.15 M NaCl, pH7.6, 0.1% (w/v) Tween®-20  
5 % (w/v) nonfat dry milk in TBS-0.1%T

**TBS-0.6%T:** 20 mM Tris, 0.15 M NaCl, pH7.6, 0.6% (w/v) Tween®-20  
1 % (w/v) nonfat dry milk in TBS-0.6%T  
10 mM imidazole in TBS-0.6%T

**STORAGE**

Refrigerate upon receipt. The product is shipped at ambient temperature.

**PROCEDURE FOR DETECTION OF POLYHISTIDINE-TAGGED PROTEINS**

Note: Volumes indicated below are for one 7 x 8.4 cm blot. Volumes may be adjusted for different sized or multiple blots.

All GoldiBlot™ reagents and other required materials should be equilibrated to room temperature prior to using for the western blotting procedure. All incubations of the GoldiBlot™ western blotting are performed at room temperature with shaking.

1. Transfer proteins from gel to a PVDF or nitrocellulose membrane.
2. Place the membrane in a tray and equilibrate with TBS-0.1%T for 3 min.
3. Block the membrane with 5 % (w/v) nonfat dry milk in TBS-0.1%T for 15 min.
4. Add 0.1 ml of GoldiBlot™ Nickel-NTA-Nanogold® to 10 mL of 1 % (w/v) nonfat dry milk in TBS-0.6%T. Vortex. Place the membrane in the solution, and incubate the blot for 30 min.
5. Wash the membrane two times with 15 mL of 10 mM imidazole in TBS-0.6%T for 2 min each.
6. Wash the membrane three times with 15 mL of deionized water for 3 min each.
7. Before starting the last deionized water wash, mix 2.5 ml GoldiBlot™ AutoMet Detect A with 2.5 mL B in a clean 15 mL container. After 5 min, add 2.5 mL C and 2.5 mL D to the mixture of A and B, and mix. Incubate the blot with 10 ml of the ABCD mixture for 6 to 15 min, or until satisfactory staining is reached.

**Note:** The incubation time of GoldiBlot™ AutoMet Detect ABCD depends on the quantities of His-tagged proteins loaded. Bands loaded with more than 100 ng His-tagged protein may be seen within 6 minutes. Longer incubation times may be needed in order to see less than 20 ng His-tagged protein. However, longer incubation may lead to some non-specific background binding.

8. Wash the membrane three times for three minutes each time with 15 mL of deionized water to terminate autometallographic amplification.

**Note:** Any light background color of the membrane back fades as the membrane dries out.

9. Air-dry the membrane.

**Note:** The concentration of NaCl and Tween 20 in binding and washes (used in GoldiBlot™ Nickel-NTA-Nanogold® binding and imidazole washes) can be slightly adjusted to achieve an optimized signal-to-noise ratio. Less NaCl and Tween®-20 can enhance the band intensity of His-tagged proteins, and higher NaCl and Tween 20 help reduce the non specific background staining.

**REFERENCES**

1. Hochuli, E.; Dobeli, H., and Schacher, A.: New metal chelate adsorbent selective for proteins and peptides containing neighbouring histidine residues. *J. Chromatograph.*, **411**, 177-184 (1987). PMID: [3443622](#). DOI: [10.1016/s0021-9673\(00\)93969-4](#).
2. Schmitt, J.; Hess, H., and Stunnenberg, H. G.: Affinity purification of histidine-tagged proteins. *Molecular Biology Reports*, **18**, 223-230 (1993). PMID: [8114690](#). DOI: [10.1007/BF01674434](#).
3. Hainfeld, J. F.; Liu, W.; Halsey, C. M. R.; Freimuth, P., and Powell, R. D.: Ni-NTA-Gold clusters target His-tagged proteins. *J. Struct. Biol.*, **127**, 185-198 (1999). PMID: [10527908](#). DOI: [10.1006/jsbi.1999.4149](#).
4. Collins, R. F.; Beis, K.; Clarke, B. R.; Ford, R. C.; Hulley, M.; Naismith, J. H.; and Whitfield, C.: Periplasmic protein-protein contacts in the inner membrane protein Wzc form a tetrameric complex required for the assembly of Escherichia coli group 1 capsules. *J. Biol. Chem.*, 281, 2144-2150 (2006). PMID: [16172129](#). PMCID: [PMC3315051](#). DOI: [10.1074/jbc.M508078200](#).
5. Wolfe, C. L.; Warrington, J. A.; Treadwell, L., and Norcum, M. T.: A three-dimensional working model of the multienzyme complex of aminoacyl-tRNA synthetases based on electron microscopic placements of tRNA and proteins. *J. Biol. Chem.*, 280, 38870-38878 (2005). PMID: [16169847](#). DOI: [10.1074/jbc.M502759200](#).
6. Bumba, L.; Tichy, M.; Dobakova, M.; Komenda, J., and Vacha, F.: Localization of the PsbH subunit in photosystem II from the Synechocystis 6803 using the His-tagged NiNTA Nanogold labeling. *J. Struct. Biol.*, 152, 28-35 (2005). PMID: [16181791](#). DOI: [10.1016/j.jsb.2005.08.001](#).